

## SECONDARY METABOLITES OF AN ALGERIAN *Phlomis bovei* AND THEIR ANTIOXIDANT ACTIVITIES

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The plants of the genus *Phlomis* are native to Turkey, North Africa, Europe, and Asia. *Phlomis bovei* De Noe, *syn. Phlomis samia* Desfontaines (Lamiaceae), is a rare Algerian endemic plant, commonly known as Kayat El Adjarah [1] in the Algerian dialect or variously named Farseouan, Tarseouan, Iniji, R'ilef, and Azaref throughout the North of Africa [2]. It is one among the nine endemic plants recorded in the Rapport National sur la Diversite Biologique [1]. *P. bovei* is a herbaceous perennial plant, which grows up to 0.8 m and often develops a stout woody base.

*Phlomis bovei* was collected from the east of Algeria in June 2006 and was taxonomically identified by Dr. Hocine Laouer from the Department of Biology, University of Setif.

About 465 g of the air powdered leaves were extracted with *n*-hexane, CH<sub>2</sub>Cl<sub>2</sub>, and MeOH in succession at room temperature. The MeOH extract was fractionated over silica gel–VLC eluting with CH<sub>2</sub>Cl<sub>2</sub> followed by increasing concentrations of MeOH. Fractions 3 and 4 (41 g), eluted with CH<sub>2</sub>Cl<sub>2</sub>–MeOH 50:50–25:75, were combined and further applied to column chromatography over silica gel using CH<sub>2</sub>Cl<sub>2</sub>–EtOAc and EtOAc–MeOH. Fraction L (1.73 g), eluted with 95:5 EtOAc–MeOH, was subjected to RP18 MPLC with H<sub>2</sub>O–MeOH. Purification of fraction L6 (18 mg) by Sephadex LH20 (MeOH) allowed the isolation of compound **1**. Purification of fraction L17 (112 mg) by SPE RP18 MeOH–H<sub>2</sub>O and TLC on silica gel (9–1) CH<sub>2</sub>Cl<sub>2</sub>–MeOH afforded compound **2**. Fraction L22 (62 mg) was subjected to SiO<sub>2</sub> TLC using (EtOAc–MeOH–H<sub>2</sub>O 100:10:5), yielding **3** (9 mg).

The structures of these compounds were elucidated by UV, <sup>1</sup>H NMR, and <sup>13</sup>C NMR, and all these data were in good agreement with the respective literature data [3–8].

**Compound 1**, C<sub>15</sub>H<sub>10</sub>O<sub>5</sub>, mp 347°C, UV (MeOH, λ<sub>max</sub>, nm): 266, 330; +NaOH: 275, 324, 390; +AlCl<sub>3</sub>: 277, 301, 341, 387; +AlCl<sub>3</sub>/HCl: 277, 301, 341, 387; +NaOAc: 278, 300, 380; +NaOAc/H<sub>3</sub>BO<sub>3</sub>: 273, 279, 350. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD, δ, ppm, J/Hz): 7.78 (2H, d, J = 8.9, H-2', H-6'), 6.90 (2H, d, J = 8.9, H-3', H-5'), 6.51 (1H, s, H-3), 6.42 (1H, d, J = 2.1, H-8), 6.22 (1H, d, J = 2.1, H-6). This compound was characterized as 5,7,4'-trihydroxyflavone (apigenin) [6].

**Compound 2**, C<sub>21</sub>H<sub>20</sub>O<sub>10</sub>, mp 226–228°C, MS–ES positive 433, UV (MeOH, λ<sub>max</sub>, nm): 269, 334; + NaOH: 267, 381; + AlCl<sub>3</sub>: 277, 349 sh, 383; + AlCl<sub>3</sub>/HCl: 279, 346 sh, 382. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD, δ, ppm, J/Hz): 7.91 (2H, d, J = 8.8, H-2', H-6'); 6.95 (2H, d, J = 8.8, H-3', H-5'); 6.83 (1H, d, J = 2.5, H-8); 6.51 (1H, s, H-3), 6.54 (1H, d, J = 2.55, H-6); 5.20 (1H, d, J = 7.5, H-1'' Glu); 3.05–3.90 (sugar protons). Identified as apigenin 7-*O*-glucoside or apigetrin [7].

Acid hydrolysis of **2** produced apigenin and D-glucose.

**Compound 3**, C<sub>10</sub>H<sub>10</sub>O<sub>2</sub>, mp 110°C, <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD, δ, ppm, J/Hz): 7.66 (2H, d, J = 8.85, H-2', H-6'); 6.73 (2H, d, J = 7.56, H-3', H-5'); 6.85 (1H, d, J = 12.6, H-4); 5.72 (1H, d, J = 12.6, H-3), <sup>13</sup>C NMR (CD<sub>3</sub>OD, 75 MHz): 112.6 (C-3), 116.7 (C-3', 5'), 126.6 (C-1'), 133.9 (C-2', 6'), 145.6 (C-4), 170 (C-2). Identified as 4-(4'-hydroxyphenyl)-*trans*-but-3-en-2-one [8].

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Compounds **1** and **2** are isolated for the first time from *P. bovei*. Compound **3** was isolated for the first time from a *Phlomis* species.

**Free Radical Scavenging Activity.** The CH<sub>2</sub>Cl<sub>2</sub> and MeOH–H<sub>2</sub>O extracts of the flowers of *P. bovei* were examined for *in vitro* antioxidant properties using the DPPH test.

The radical-scavenging activity was measured by the molar ratio of antioxidant to DPPH radical required for 50% reduction in DPPH radical concentration in 45 min. The results of our experiments demonstrated that the two compound-extracts tested possess radical-scavenging activity. It was also found that the free-radical-scavenging activity of MeOH–H<sub>2</sub>O extract (EC<sub>50</sub> 0.30) was stronger than that of CH<sub>2</sub>Cl<sub>2</sub> extract (EC<sub>50</sub> 0.50).

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